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Polymyxin conjugates.

@ Water-soluble polymyxin-carrier conjugates such as polymyxin B-dextran conjugates are disclosed. They have a greater half-life in the bloodstream and are significantly more potent and less toxic than native polymyxins.

POLYMYXIN CONJUGATES

The invention relates to novel polymyxin conjugates and to methods of preparing and using them.

Endotoxins or lipopolysaccharides are structural molecules derived from the cell walls of the Gramnegative basteta. When introduced into the bloodstream, they can interfere with the regulation of body temperature and cause lever. They also have a twice effect, leading to cardiac, pulmonary and kiney failure. Endotoxin-related diseases are a leading cause of death among those patients in intensive care units.

Unique among artibiotics is the ability of polymyxins, especially of polymyxin B (PMB) to neutralize endotoxin, accomplished by biholing to the lipid A region of the endotoxin molecule. Polymyxin B from Bacillus polymyxa (8. aerosporus) is a highly charged amphiphilit cyclic peptidolipid. It is also useful in combating various fungal infections, especially those arising in immunocompromised individuals.

However, PMB has some properties which renders it less than an ideal antibiotic. First, it has a short half-life in the body, requiring repeated closeges in order to be effective. Secondly, as it passes through the kidney it can cause extensive damage. Thirdly, at high doses it possesses neurotoxic properties which cause respiratory paralysis.

Previously, researchers have conjugated polymyxin to immobile or fixed molecules. See e.g. Issekutz, J. Immunol. Methods 81 (1983) 275-281, describing the binding of PMB to Sopharose. These conjugates, while useful in purification techniques, are not suitable for in vivo therapeutic use.

One approach to achieve pharmacological activity, increased duration, or decreased organ toxicity has involved the conjugation of drugs to large molecular weight macromolecules such as dextran, polyethylene or developed the conjugation of drugs to large molecular weight macromolecules such as dextran, polyethylene organization of the conjugation have been met with only limited success, however. For example, the conjugated form of procalemide (an antiarrythmic drug) was less active and exhibited a shorter half-life than native procalemide (Schacht et al., Ann. NY Acad. Sci. (1985) 448 199-211). Similarly, a prostaplandin analog, 8245, linked to a carrier, was less effective (by several log orders) than the native molecule (Bamford et al., Bioch. Biophys. Acta 886 (1989) 199-118). Reductions in 50 biological potency have also been described for conjugated forms of kallifran, parollinin, bradykinin (Odya et al., Biochem. Pharmacol. 27 (1979) 173-179] and the anti-tumor drugs deunorubicin (Hurritz et al., Appl. Biochem. Patrancol. 27 (1979) 173-179] and the anti-tumor drugs deunorubicin (Hurritz et al., J. Appl. Biochem. 2 (1980) 25-35) and mitomycin (Takakura et al., Cancer Res. 44 (1984) 2505-2510; Conjugated enzymes also suffer a reduction in biological activity due to Staric Indirance and reduction in biological activity due to Staric Indirance and reduction in biological activity due to Staric Indirance and reduction in biological activity due to Staric Indirance and reduction in biological activity due to Staric Indirance and reduction in biological activity due to Staric Indirance and reduction in biological activity due to Staric Indirance and reduction in biological activity due to Staric Indirance and reduction in biological activity due to Staric Indirance and reduction in biological activity due to Staric Indirance and reduction in biological activity due to Staric Indirance and reduction in biological activity due to Staric Indirance and reduction in biological activity due to Staric Indirance and redu

It would be desirable to develop a form of polymyxin which would stay in the blood stream longer and/or does not have neuro- or nephrotox/city at therapeutic doses.

The present invention relates to polymyxin-carrier conjugates which are soluble in water, e.g. polymyin A. B- or E-carrier conjugates, particularly PMB-carrier conjugates. They are indicated for use in neutralizing endotoxin. They are an improvement over the administration of native polymyxin because they are more effective and are less toxic.

As used herein the following definitions apply:

- 40 LD₀ is the highest non-toxic dose of PMB or its conjugate;
 - LO₁₀₀ is the dose of PMB or its conjugated form which results in 90-100% lethality when injected into a normal test animal;
 - PD₁₀₀ is the dose of polymyxin B or its conjugated form which, when injected into a hypersensitized test animal, results in at least 95 % survival;
- 45 Therapeutic Index (TI) is calculated by dividing LD100 by PD100.

As defined herein a conjugate is water-soluble when it has a solubility in water of at least 0.5 mg/ml at 25° C, preferably of at least 1 mg/ml at 25° C. Such conjugates are water-soluble and injectable, non-matrix, colloid, not cross-linked, non-macromolecular, and/or non-particulate.

The carrier which can be conjugated to polymyxin is chosen from molecules which can form watersoluble conjugates and which are uniform, non-toxic, non-carcinogenic, non-instituting and non-immunogenic. It is normally a biopolymer. Such carriers include polysecharides such as dextran or hydroxy ethyl starck (HES), proteins such as elbumin, and polymers such as polyvinyl pyrrollidone, polyethylene glycol and polyvinyl alcohol. Dextran is the most preferred.

The size of the blopplymer portion, i.e. the carrier part of the conjugate may vary. Typically it will range from a molecular weight of 25,000 to 500,000, preferably from 50,000 to 300,000, more preferably from

79,000 to 200,000. The size of the biopolymer chosen can significantly contribute to the duration of the conjugate's effective time of circulation in the blood stream. Generally, the larger the biopolymer, the longer the conjugate will stay in the circulation. Thus the size of the biopolymer can be adjusted to result in a conjugate whose time of duration in the body corresponds to a predetermined time.

The conjugates of the invention may be prepared in conventional manner.

Taking polymyxin B as an illustration, this has five -pamino groups which bind to the bacterial endotoxin. At least one -pamino group can be used to securely bind the PMB to the carrier, but all five sites cannot be used for this purpose, or the conjugate will lack endotoxin-neutralizing activity. Various chemical reactions can be used to conjugate polymyxin to these carriers. The number of PMB molecules bound per nolecule of polymer can be influenced by varying the ratios of reactants during the coupling reaction. Generally the conjugate will contain between 1 PMB: 1 biopolymer to 200 PMB: 1 biopolymer, more repreferably the conjugate will have fron 1-10 PMB: 1 biopolymer, and even more preferably from 1-3 PMB: 1 biopolymer and the public production of the properties of the

One method of making a polymyxin-carrier conjugate, more specifically, a PMB-dextran conjugate, is through carbamate linkage (method A). PMB-dextran conjugates made in this fashion (detailed in Example 1, below) are found to retain the anti-endotoxin activity of native PMB, and in addition are found to be devoid of the acute neurotoxicity exhibited by native PMB. These conjugated forms also show a 2-5 fold improved therapeutic index by decreasing chronic toxicity over that of native PMB.

20 A second method of making conjugated polymrykin dextran is through an amine bond (method B). PMB-dextran conjugates made in this manner (detailed in Example 2, below) retain the same general anti-endotoxin activity or are more active than native PMB. These conjugated forms are completely devoid of any acute neurotoxicity seen by native PMB and exhibit a 33-fold improvement of the therapeutic index by reducing chronic toxicity over that seen with native PMB. In particular, now PMB-dextran conjugate is devoid of any measurable toxicity at the doses tested and retains good anti-endotoxin activity, resulting in an 80-fold improvement in the therapeutic index.

Further, It has been surprisingly found that conjugates produced by this method and "rapidly purified" exhibit a 60- to 120-fold improvement in TL "Rapidly purified" as used throughout the specification and claims means that the material is purified from unbound PMB using molecular sleving chromatography, desalting gels, or Amicon ultrafiltration, occurs for approximately 12-24 hours, preferably approximately 18 hours (as opposed to an extended dialysis time of 7-10 days). Thus conjugates produced by these methods and rapidly purified comprise another and preferred aspect of this invention.

The polymydn-carrier conjugate obtained from either of the two methods outlined above is, however, difficult to purify initially, a 10-day extended dalaysis was tried in corte to separate nather PMB from the conjugate. Although the protein level inside the dialysis bag reached an asymptote, and gel permeation column chromatography showed a single large molecular weight species, the material still contained unbound PMB. Even when pressure dialysis (Amicon) filtration was substituted for extended dialysis the results were the same. Conjugates purified in this manner were equipotent to PMB, but still retained 1/3 to 1/5 the tocklicy of PMB. While not intending to be bound by theory, it appears that there is some chemicate association between the "free" PMB and the conjugate which renders its separation difficult. Upon injection in the animal the "free" PMB dissociates itself and causes toxicity problemant the "free" PMB dissociates itself and causes toxicity problemant.

The following procedure (method C) (detailed in Exemple 4, below) obviates the above difficulties. Polymykin-dextran conjugates are precipitated in a lower alcohol, preferably methanol, followed by centrifugal collection and ultrasonic resolubilization. This procedure is repeated, normally at least time times, and preferably 7 to 10 times and the resulting material is evaluated for purity, e.g. by gel permeation chromatography and reverse-phase high pressure liquid chromatography (RP-HPLC). If protein is detected by either method, the precipitation-resolubilization procedure is repeated. Conjugate purified in this manner is referred to herein as "ultra-pure" polymyxin-conjugate and generally can be described as having less than 20 µg unconjugated polymyxin per 1 mg total protein, and preferably sess than 10 µg unconjugated polymyxin per 1 mg total protein.

A further aspect of the invention is thus a process for the preparation of a water-soluble polymyxindextran conjugate comprising

- a) appropriately conjugating polymyxin to dextran to form a water-soluble polymyxin-dextran conjugate;
- b) precipitating the resultant conjugate in a lower alcohol;
- c) resuspending the conjugate; and
 - d) repeating steps b) and c) until the conjugate is substantially free from unbound polymyxin.

"Substantially free from unbound polymyxin" is herewith defined as a ratio of conjugate to unbound polymyxin of at least 95:1, preferably of at least 99:1 on a molar basis. The lower alcohol is e.g. of 1 to

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10 carbon atoms, it preferably is of 1 to 4 carbon atoms, e.g. n-butanol or isopropanol, more preferably methanol or ethanol. The polymyxin preferably is PMB.

Ultra-pure conjugate has virtually no toxicity and as a result an LD tee cannot be established for it. At a dose of 100 mg/kg no toxicity is seen. At higher concentrations the solution is too viscous to inject. This is in contrast to native PMB which has an LDs i.v. of 5 mg/kg. Thus the Therapeutic Index of ultra-pure PMB conjugate is over 1000-fold higher than native PMB, due to decreased toxicity.

A second advantage in using the conjugate rather than native polymyxin is that the conjugate has a longer duration of activity and thus can be used prophylactically. Thus one espect of the Invention is the prevention of diseases caused by the presence of bacterial endotoxin by administering to a subject in the odd of such prophylactic treatment a prophylactically effective amount of a water-soluble conjugate of polymyxin, preferably PMB and a carrier, particularly dextran, and a pharmacoutically acceptable further carrier or dilluent as appropriate, e.g. an inent solution. As the conjugate has virtually no toxicity it can be prophylactically administered on a routine basis to patients who might be susceptible to septic shock, such as those dependent upon floatif foot for fonce-term quorityment.

The invention also concerns a pharmaceutical composition comprising a water-soluble polymyxin-carrier conjugate togethor with further pharmaceutically acceptable carriers or diluents as appropriate, e.g. an inert solution.

The invention also concerns a process for the preparation of a pharmaceutical composition comprising mixing a water-soluble polymyxin-carrier conjugate with further pharmaceutically acceptable carriers or diluents as exponentiate, e.g. an Inert solution.

Another aspect of the Invention is the treatment of diseases caused by the presence of bacterial andctoxin comprising administering to a subject in need of such treatment an endotoxin-neutralizing amount of a water-soluble conjugate of polymyxin, particularly PMB and a carrier, particularly dextran, and an inert as solution. As polymyxin is also effective in combating bacterial or fungal Infections, another sepect of this invention is a method of treating bacterial or fungal infections comprising administering to a subject the of such treatment a bacteriocidally- or fungicidally-effective amount of a water-soluble conjugate of polymyxin and a carrier, particularly dextran, and an inert solution.

The polymyxin-carrier conjugate of the Invention can be used in a manner consistent with the use of polymyxin itself, i.e. it can be used alone as an antibiotic for bacterial or fungal infactions or combined with other bacteriocidal agents and/or anti-infammatory agents, it may be administered, e.g. Intramuscularly, intravenously, intraflecally, acconjunctivally or topically. Thus formulations for intramuscular injections typically comprise an effective amount of PMB-conjugate in startie water, physiological saline or approximately 1 % process* HCI. Intravenous formulations typically comprise an effective amount of PMB-conjugate in 5 % dextrose and startie water. Intraflaced informulations typically comprise an effective amount of PMB-conjugate in physiologic saline. For topical ophthalmic use, an effective amount can be mixed with water or physiological saline, and optionally glycenine, and cupric sulfate for eye drops, or it may be made into an orithment or suspension. Creams, for topical applications, especially for burned areas, typical into an orithment or suspension. Creams, for topical applications, especially for burned areas, typical proprise an effective amount of polymyxin-conjugate in a base of inscrive ingredients such as liquid periolatur, propylene glyco, polyoxyethylene, polyxycyproplene and emulsifyling wax.

The amount of polymyxin-conjugate to be used is based on the amount of native polymyxin which would typically be prescribed for a particular patient (taking into account such factors as the condition being treated and age and weight of the patient) and the activity of the particular polymyxin-conjugate used. Often, a disage reduction of up to 50 % compared to native polymyxin can be realized due to the effective increased duritivity of the polymyxin conjugate, its reduced toxidity and its increased duration of activity.

The invention is illustrated by reference to the following non-limiting Examples. All temperatures are in degrees Centigrade.

Example 1: Chemical conjugation of PMB to dextran

(Method A; carbamate linkage)

5 2 g dextran (79,000 or 200,000 MW) is dissolved in 20 ml water, cooled to 0°, and 5-300 mg of 1-cyano-4-dimethylamino pyridinium tetrafluoroborate (CDAP) is added and mixed for 30 seconds. Triethylamine (0.2 M, 0.04 ml per 5 mg CDAP) is added dropwise with vigorous stirring, and the entire reaction mixture is transferred to 80 ml of ice cold absolute ethanol containing 1 ml of 10 N HCI. The

destran precipitates, and the precipitate is removed by centrifugation at 1000 x g for 5 min at 0°, and resolubilized in 29.50 ml 0 0.25 M Na-bicarbonate buffer at pH 9.0. To this mixture 600-1000 mg of PMB (either powdered or presolubilized in water) is added and stirred for 24 hours at 8°. The entire reaction mixture is then transferred to a 50,000 molecular weight cut-off dialysis tubing and dialyzed against 0.05 M s pyrogen-free phosphate buffer for 6°10 days. The final dialyzed reaction mixture is measured for protein content by spectrophotometry at 208 nm absorbance or at 595 nm using the Bradford reagent (Bio Rad, Richmond, CA, USA).

Free amino groups are determined using the ninhydrin reaction, with native PMB as a control. Analysis for nitrogen and carbon content is done using a CHN Elemental Analyzer.

In the table below, "molar ratio" is determined by dividing the concentration of dextran used in conjugation (23.7 µmol) by the final PMB-protein concentration in µmol (based on 208 nm analysis) after dialysis. Dextran is abbreviated "dx". The "C.N" ratio assumes 450 molecules of water per 2 g dextran. The "bondsPMB" is an estimate of the number of bonds by which PMB is attached to dextran, based on the nithydrin reaction. "ND" is 'not determined."

Summary of conjugation reactions				
Reaction	CDAP	molar ratio	C:N ratio	bond/PMB
A	21 µmol	1 dx: 0.23 PMB	ND	ND
В	106 µmol	1 dx: 1.62 PMB	1 dx : 1.52 PMB	3.0
BB	106 µmol	1 dx: 1.95 PMB	1 dx: 1.90 PMB	1.1
С	532 µmol	1 dx : 3.70 PMB	1 dx: 3.25 PMB	3.0
cc	532 µmol	1 dx:4.99 PMB	1 dx:5.00 PMB	2.2
D	1064 umol	1 dx: 6.62 PMB	1 dx:8.70 PMB	1.7
DD	1064 µmol	1 dx : 8.81 PMB	ND	1.3
E	2128 µmol	1 dx :10.9 PMB	ND	2.1

Example 2: Chemical conjugation of PMB to dextran

35 (Method B; smine bond)

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1.25 g dextran (79,000 or 200,000 MW) is dissolved in 20 ml of distilled water and than 0,071-0,71 g Na-periodate is added. After 1 hour at 22° the reaction mixture is transferred to a column containing DEAE A25 cationic exchange resin (Pharmacia Inc., Piscataway, NJ, USA) and the mixture is collected and pooled to a single fraction of 20-25 ml. The oxidized dextran is then mixed with 2 g PMB dissolved in 80-250 ml Na-bicarbonate or Na-borate buffer (pit 8.5-90) and atter 60 minutes, 40 ml 0.05 - 25 % Na-borohydride either in a single treatment or in multiple treatments, each treatment leating 30 minutes to 24 hours is added. This reaction proceeds for 30 minutes and then is dialyzed for 7-10 days at 80 against 0.01 minutes and then is dialyzed for 7-10 days at 80 against 0.01 minutes and then is dialyzed for 7-10 days at 80 against 0.01 minutes and then is dialyzed for 7-10 days at 80 against 0.01 minutes and then for 30 minutes and then or 7-10 days at 80 against 0.01 minutes and then is dialyzed for 7-10 days at 80 against 0.01 minutes and then is dialyzed for 7-10 days at 80 against 0.01 minutes and then is dialyzed for 7-10 days at 80 against 0.01 minutes and then or 7-10 days at 80 against 0.01 minutes and then is dialyzed for 7-10 days at 80 against 0.01 minutes and then or 7-10 days at 80 against 0.01 minutes and 10 minutes 10 minutes

In addition to purification by extended dialysis, several representative samples are rapidly purified using dialysis for 18 hours, purified on a G-100 Sephadex column (Pharmacla, inc., Piscataway, NJ, USA) and concentrated on an Amicon filtration unit using a YM-100 Amicon filter.

In the table below the motar ratio is determined by dividing the amount of dextren used in conjugation (6.25 µmol) by the µmol final PMB protein (based on the 208 nm data) after dialysis. The percent nitrogen is determined using a CHN elemental analyzer. BondsrPMB is estimated by the ninhydrin reaction.

Summary of conjugation reactions				
Reaction	NaIO ₄ (g)	molar ratio	% nitrogen	bonds/PMB
1/50	0.014	1 dx : 3.64 PMB	0.21	3.6
1/10	0.071	1 dx ; 12.9 PMB	1.05	2.9
1/5	0.142	1 dx: 26.8 PMB	1.71	2.6
1/4	0.178	1 dx:29.8 PMB	2.76	3.0
1/21	0.355	1 dx: 39.6 PMB	3.74	2.2
stock*	0.710	1 dx :151 PMB	6.91	4.0

^{*}Reaction mixture is turbid and remains so after dislysis

Example 3: Anti-endotoxin activity of PMB-conjugates

A. Endotoxin-Induced (ethality

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Male CB57BL/c mice (16-22 g, Jackson Labs) are used throughout this study. Animals are given endotoxin (01116. from Lst Biologials, Palo Alto, CA, USA) and galactosamine to hypersensitize, as described in Galanos et al., Proc. Natl. Acad. Sci. USA 76 (1979) 5939. The endotoxin and galactosamine (320 mg/kg) are injected intraperitoneally in 0.5 ml pyrogen-free isotonic saline per animal between 13:00 and 15:00 to avoid clumal variation. From a dose-response study, it is determined that 0.01 mg/kg endotoxin produces 85:45 % lethality. Animals ares-response study, it is determined that 0.01 mg/kg endotoxin produces 85:45 % lethality. Animals ares-response study for 6 days after injections.

B. PMB (native and conjugated forms) induced lethality

PMB (native and conjugated forms) are evaluated for their inherent toxicity in non-sensitized mice. Male CB57BL/c mice (18-22 g, Jackson Labs) are used throughout this study. Animals are given either native PMB or conjugated PMB by intrapertioneal injection (0.5 m/lanimal) and lethality is monitored for 7 days. The LD₁₀₀ is determined by varying the dose of PMB (native and conjugated forms) that result in 100 % lethality. Form 10 to 20 animals are used for each dose.

C. PMB anti-endotoxin evaluations

PNB (native and conjugated forms) are evaluated for their ability to neutralize endotoxin by either premixing with endotoxin for 60 minutes before i.b. injection or by locadministration of the substances as a single i.b. injection. While the concentration of the PMB (native and conjugated forms) are varied, the volume of each injection is kept constant at 0.5 ml per animal. The protection by the conjugated PMB is compared to the native PMB (in terms of mg protein/kg). Controls (endotoxin, galactosamine and vehicle) are included as well in each study. Each group has between 10 and 17 animals.

Statistical analyses of survival between groups or in relation to controls is performed using a Chi-square analysis using the Yate's correction for continuity.

D. Determination of Therapeutic Index

The Therapeutic Index (Ti) for PMB (native and conjugated forms) is determined by dividing the LD₁₀₀ by the PD₁₀₀. In the table below, the "treatment" refers to the conjugates appearing in Examples 1 and 2, supra.

^{*}Purified using rapid dialysis

Anti-endotoxin data			
Treatment	LD ₁₀₀ (mg/kg)	PD ₁₀₀ (mg/kg)	TI
Native PMB ¹	25	2	12.5
Native PMB ²	25	2	12.5
A	>30	1	>30
В	>28	>2	>14
С	ND	5	ND
D	140	. 2	70
1/5*	83	0.2	415
1/21*	about 40-60	0.05	800-1200
stock ²	>212	0.2	>1060
stock ³	>212	20	>10.6

¹Rapidly-purified material

Example 4: Ultra-pure PMB-dextran conjugate

(Method C; alcohol precipitation)

Preparation

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PMB-dextran conjugate is prepared essentially as described in Example 2, supra, to obtain conjugate designated "RXN 1/50".

The conjugets, a white flocculant, is precipitated in 60 % methanol, collected by centrifugation, and then subjected to ultrasonic resolubilization. This process is repeated seven times and the resulting material is evaluated for purity by gol permeation chromatography and reverse phase high pressure liquid chromatography (RP-HPLC). Although no free PMB is detected by gel permeation chromatography, RP-HPLC shows 48 µg unconjugated PMB per mg of total protein. The conjugate is then subjected to 3 additional precipitations to remove the unconjugated PMB.

2. Biological activity

The resulting ultra-pure PMB-dextran conjugate is compared to native PMB in an endotoxin-induced lethality test as described in Example 3, C., supra.

At 1 mg/kg [iv. the ultra-pure PHI-conjugate gives > 70 % protection against a LD₅₀ endotoxin challenge for at least 6 hours. At 10 mg/kg iv. this conjugate provides therapeutic protection 1.5 hours after administration of the endotoxin. Therefore the PMB-conjugate is suitable for prophylacitic use.

The ultra-pure PMB-conjugate has a surprisingly reduced toxicity as compared with native PMB. Native PMB has an LDo. i.v. of 5 mg/kg and an LDr.o. i.v. of 9.5 mg/kg, suggesting an early onset of toxicity and narrow therapeutic range. The ultra-pure PMB-conjugate has an LD, of > 100 mg/kg and it has not been possible to establish an LDr.o. due to its Inhorent non-toxicity. Comparisons in the table below are made between the highest non-toxic dose of PMB (native or conjugated):

²The PMB or conjugate is allowed to react with the endotoxin for 60 min before i.p. injection

³The PMB or conjugate is injected simultaneously with the endotoxin The fractions designate reaction conditions based on the amount of sodium periodate used in the conjugation reaction. If 710 mg is multiplied by the fraction the amount (in mg) of sodium periodate used in each sample is obtained.

Biological activity			
Treatment	LD ₀ (mg/kg) ¹	PD ₁₀₀ (mg/kg) ²	TI
FMB ultra-pure conjugate	5 > 100	0.1 0.1	50 > 1000

¹ PMB (native or conjugate) given by i.v. injection and lethality monitored

² PMB (native or conjugate) given by i.v. injection 1-3 minutes before an LD₉₀ l.p. Injection of E.coli 0111B4 endotoxin (0.5 μg/kg)

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- 1. A water-soluble conjugate of a polymyxin and a carrier.
- 2. A conjugate according to claim 1 wherein the carrier is a polysaccharide such as dextran or hydroxyethyl starch, a protein such as albumin, or a polymer such as polyvinylpyrrolidone, polyethylene glycol or polyvinyl alcohol.
 - 3. A conjugate according to claim 1 wherein the polymyxin is PMB.
 - 4. A conjugate according to claim 2 wherein the carrier is dextran.
 - 5. A conjugate according to claim 3 wherein the carrier is dextran.
 - A conjugate according to claim 5 wherein PMB is attached through carbamate linkage. 7. A conjugate according to claim 5 wherein PMB is attached through amine bonds.

 - 8. A process for the preparation of a polymyxin-dextran conjugate according to claim 4 comprising:
 - a) appropriately conjugating polymyxin to dextran to form a water-soluble polymyxin-dextran conjugate: b) precipitating the resultant conjugate in a lower alcohol;
 - c) resuspending the conjugate; and
 - d) repeating steps b) and c) until the conjugate is substantially free from unbound polymyxin.
 - 9. A pharmaceutical composition comprising a conjugate according to claim 1 together with further pharmaceutically acceptable carriers or diluents.
- A conjugate according to claim 1 for use as a pharmaceutical.

Claims for the following Contracting States: ES.GR

- 1. A process for the preparation of a pharmaceutical composition comprising mixing a water-soluble conjugate of a polymyxin and a carrier together with further pharmaceutically acceptable carriers or diluents.
- 2. A process according to claim 1 wherein the carrier is a polysaccharide such as dextran or hydroxyethyl starch, a protein such as albumin, or a polymer such as polyvinylpyrrollidone, polyethylene glycol or
- 3. A process according to claim 1 wherein the polymyxin is PMB.
- 4. A process according to claim 2 wherein the carrier is dextran.
 - 5. A process according to claim 3 wherein the carrier is dextran.
 - A process for the preparation of a water-soluble polymyxin-dextran conjugate comprising:
 - a) appropriately conjugating polymyxin to dextran to form a water-soluble polymyxin-dextran conjugate;
 - b) precipitating the resultant conjugate in a lower alcohol:
 - c) resuspending the conjugate; and
 - d) repeating steps b) and c) until the conjugate is substantially free from unbound polymyxin.
 - A process according to claim 6 wherein the conjugate is a PMB-dextran conjugate.
- 8. A process according to claim 6 wherein steps b) and c) are repeated at least three times.



EUROPEAN SEARCH REPORT

Application Number

	DOCUMENTS CONS	INFERENCE OF BELLEVIA	NPP	EP 90 81 08
		IDERED TO BE RELEVA indication, where appropriate.	Rejevant	CLASSIFICATION OF THE
Category	of relevant		to claim	APPLICATION (Int. Cl. 5)
X	line 9; page 11, 1	page 5, line 2; page , line 30 - page 7, ine 31 - page 13, example 1; page 27.	1-10	A 61 K 47/48
х	residual colistin	o. 85:421296, ogical abstract AWA et al.: immunoassay of pp](cation to detect	1-3	
Y	IDEM		1-10	
Y	EP-A-0 147 761 (M INC.) * Page 1, lines 4-		1-10	TECHNICAL FIELDS SEARCHED (Int. CLS)
	The present search report has	been drawn up for all claims	-	
	Place of search HAGUE	Date of completion of the search		Examiner

EPO FORM ISO OCAT (PRO) CATEGORY OF CITED DOCUMENTS

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 after the filing date
 D: document cited in the application
 L: document cited for other reasons

- & : member of the same patent family, corresponding document